

Fast Hydrogen Peroxide Assay

Fluorometric and Colorimetric Assay

Catalog Number

TBS2066-100

TBS2066-200

Kit Size

100 assays

200 assays

DESCRIPTION

Hydrogen Peroxide (H_2O_2) is a reactive oxygen metabolic byproduct that serves as a key regulator for a number of oxidative stress-related states. It is involved in many pathological processes of diseases such as asthma, inflammatory arthritis, atherosclerosis, diabetic vasculopathy, osteoporosis, neuro-degenerative diseases, Down's syndrome, and immune system diseases.

Tribioscience's ADHP-red Hydrogen Peroxide Assay Kit is a sensitive, simple, direct, and HTS-ready colorimetric/fluorometric assay designed to measure H_2O_2 levels in biological samples. In the presence of Horse Radish Peroxidase (HRP), the ADHP-red probe reacts with H_2O_2 to produce a resorufin that appears red in color. The resorufin can be measured with absorption of 570nm ($\lambda_{max} = 570nm$) for colorimetric or a highly stable fluorometric assay at maximum emission of 585nm (Ex/Em = 535/587 nm).

APPLICATION

- Detection of HRP activity on liquid systems such as ELISA.
- Western Blot or other immunoassays.
- Custom packaging and bulk purchase information are available upon request

FEATURES

- F**lexible: Suitable for colorimetric and fluorometric methods.
- A**ccurate: Use 50 μ L samples. Detection ranges 0.4-200 μ M in 96-well plate for colorimetric assay and fluorometric assay.
- S**imple and high-throughput: One-step procedure: just load-incubate-Read. The kit can be used for a robust method.
- T**ime-saving less than 30 minutes

KIT CONTENTS

Component	100x Rxns	200x Rxns
ADHP solution	60 μ L	120 μ L
H_2O_2 Standard (0.88M)	50 μ L	100 μ L
HRP Reagent	100 μ L	200 μ L
1x Reaction buffer	10ml	20mL

Storage conditions: Store the kit at $-20^{\circ}C$ protected from light.
Shelf life: 6 months.

ASSAY PROCEDURES

1. Sample Preparation

Collect cell culture supernatant, serum, plasma, urine and other biological fluids. Centrifuge for 5-minutes at 5000 xg then collect supernatant for analysis. It is recommended with all sample types to assay immediately or aliquot and store the samples at $-80^{\circ}C$. Avoid repeated freeze-thaw cycles. Add 50 μ L samples into each well, then bring the volume to 100 μ L with assay buffer.

2. Standard Preparation:

2.1 Dilute 2.0 μ L of 0.88M H_2O_2 into 878 μ L of 1x Assay buffer to generate a 2mM H_2O_2 stock solution (*Note: 2 mM H_2O_2 stock*

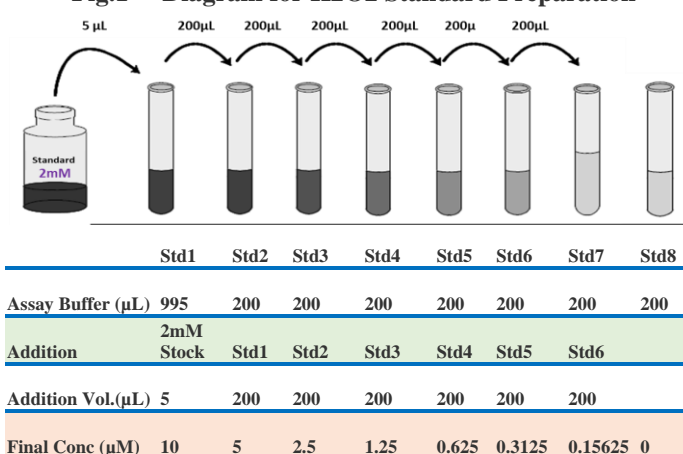
solution prepared in this step will be less stable and should be used within a few hours of preparation, although the H_2O_2 stock solution (0.88M) has been stabilized to slow its degradation).

2.2 Label 8x 1.5mL tubes 1-8 for a standard curve as shown in the diagram below.

Add 995 μ L of 1x Assay Buffer to Std 1 and 200 μ L to Std 2-8.

2.3 Add 5 μ L of 2mM H_2O_2 Stock solution to Std1 then transfer 200 μ L of Std1 to Std2. Carry out a 2x serial dilution for Std 3-7. Leave Std 8 as the 0 standard (the assay buffer alone). The standards concentrations are 10, 5, 2.5, 1.25, 0.625, 0.3125, 0.15625, and 0 μ M for Std 1-8.

Fig.1 Diagram for H_2O_2 Standard Preparation



2.4 Load the samples: Pipet 50 μ L of standards, controls, and test samples into individual wells of a microplate in duplicate manner (*Note: recommend to run a pilot study to determine the optimal concentration of sample within the assay standard curve range*).

2.5 Prepare ADHP-red working solution by mixing the reagent as following:

- 50 μ L ADHP-red reagent stock solution
- 100 μ L HRP stock solution
- 4.85mL of 1x Reaction Buffer

This 5mL volume is sufficient for ~100 assays. Note that the final concentration of each component will be two-fold lower in the final reaction volume.

2.6 Begin the reactions. Add 50 μ L of above ADHP-red working solution to each microplate well containing the standards, controls, and samples.

2.7 Incubate the reactions. Incubate at room temperature for 30-minutes protected from light. Because the assay is continuous (not terminated), you may measure fluorescence or absorbance at multiple time points to follow the kinetics of the reactions.

2.8 Measure fluorescence or absorbance using a microplate reader with excitation range of 530-560 nm and fluorescence

emission detection at ~590nm, or absorbance at ~560nm.
2.9 Correct for background fluorescence or absorbance. For each point, subtract the value derived from the no-H₂O₂ control.

Calculation

Subtract the blank value (0μM Standard) from the standard values and plot the ΔOD or ΔF against standard concentrations.

Determine the slope and calculate the glucose concentration of the Sample using the equation obtained from the linear regression of the standard curve. $H_2O_2 = N \times (R_{\text{sample}} - R_{\text{blank}}) / \text{Slope} (\mu\text{M})$
Where: R_{sample} and R_{blank} are optical density or fluorescence intensity readings of the sample and blank, respectively. N is the sample dilution factor.

The Typical data is displayed in Fig.2 and Fig 3.

RELATED PRODUCTS:

ATP Activity Assay (TBS2010)
ADHP Red Hydrogen Peroxidase Assay Kit (TBS2067)
AmplexRed_HRP-System (TBS5026)
Tryptase Activity Assay (TBS2101)
β-Hexosaminidase Activity Assay (TBS2105)
Cytochrome C Oxidase Activity Assay (TBS2115)
Fast Glucose Determination Colorimetric/Fluorometric Assay (TBS2087)
Glucose Oxidase Activity Colorimetric/Fluorometric Assay (TBS2088)
Non-esterified Fatty Acid Assay (TBS2203)
Glycerol Colorimetric / Fluorometric Assay (TBS2204)
Protein Assay Kits (TBS2005)
Cell Nuclear Extract kit (TBS6025)

For research use only.

