

DESCRIPTION

EasyCloning Kit is designed for quick and effective PCR cloning. With our proprietary recombinase enzyme, this kit is especially powerful in directly cloning any single or multiple PCR products into destined vector effectively without the tedious and limiting tasks such as selecting proper restriction enzymes, phosphatases, or ligases.

KEY FEATURES

- **Fast:** One step cloning procedure completed within 30 minutes.
- **Easy:** PCR DNA and linearized vector immediately used in cloning, no restriction enzymes, phosphatases or ligases are needed.
- **Effective:** Long PCR DNA up to 100 kb can be efficiently cloned vectors with successful rate >95%.

APPLICATIONS

- PCR cloning up to 100 kb
- Gene transfer from one vector to another
- High-throughput (HTP) PCR cloning
- *In vitro* joining of DNA fragments
- Gene insertion, knockout, mutagenesis et.al

CONTENTS for 10 or 20 Reactions

Kit Components	TBS4050-10	TBS4050-20
EasyCloning Enzyme (5 U/μl)	10 μl	20 μl
10X CloneEZ® Buffer	20 μl	40 μl
Manual	1	1

STORAGE

The kit should be stored at -20°C. It will remain stable for at least one year.

GENERAL PROTOCOL

A. PCR Amplification of Target DNA: To clone any DNA fragment into a linearized vector using this kit, the insert fragment should be obtained by PCR using primers with an add-on of 15-30 base sequences homologous to either side of the restriction site that is used to linearize the vector (longer than 15 base will increase efficacy but no more increase after 30 bases). Therefore, both primers should cover a 15-30 base

Water bath or heating block (42°C)

SOC liquid medium

DH5α competent cells (>1×10⁸ cfu/μg)

1. Thaw one vial of frozen 50 μl competent cells on ice. Tap the tube gently to ensure that the cells are suspended.
2. Add 5–8 μl of reaction mixture to the competent cells. Tap the tube gently and incubate the tube on ice for 30 minutes.
3. Heat shock the cells by placing them in 42°C water bath for 45–90 seconds and then place the tube on ice for 2–3 minutes.
4. Add 600 μl of SOC medium to the cells and then incubate the cells on a shaker set at 250 rpm at 37°C for 60 minutes.
5. Centrifuge the cell down at 4000 rpm for five minutes and then remove and discard about 500 μl of medium. Gently suspend the cells by tapping the tube.
6. Transfer 10 μl and 100 μl of the suspension to two different plates containing appropriate antibiotics, respectively. Spread the cells evenly on the plates.
7. Incubate the plates overnight at 37 °C.

TROUBLESHOOTING

Problem	Probable Cause	Solution
Few or no colonies are obtained from the transformation.	The competent cells have low transformation efficiency.	Check the transformation efficiency. Competent cells with >1×10 ⁸ cfu/μg are recommended.
	Too much reaction mixture is used.	Do not add more than 10 μl of reaction mixture to 50 μl of competent cells. Too much reaction mixture inhibits the transformation.
	There are inhibitory contaminants from PCR DNA or from linearized vector.	Both of the PCR DNA and the linearized vector should be purified.
	The molar ratio of vector to insert is off.	Usually an insert/vector molar ratio of 2:1 is optimal. If the insert is as large as the linearized vector, a molar ratio of 1:1 can also be used.
Most of the colonies contain no insert.	The cloning vector is not completely linearized.	Gel-purify the linearized vector.
	The cloning reaction is contaminated with plasmids having the same antibiotic resistance.	Purified PCR DNA may contain the template plasmid, so gel-purify the PCR DNA.

Note: even there are only one or two colonies, it is likely to be positive clones.

This product is for Research Use Only.

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